The Open Access Israeli Journal of Aquaculture – Bamidgeh

As from **January 2010** The Israeli Journal of Aquaculture - Bamidgeh (IJA) will be published exclusively as **an on-line Open Access (OA)** quarterly accessible by all AquacultureHub (<u>http://www.aquaculturehub.org</u>) members and registered individuals and institutions. Please visit our website (<u>http://siamb.org.il</u>) for free registration form, further information and instructions.

This transformation from a subscription printed version to an on-line OA journal, aims at supporting the concept that scientific peer-reviewed publications should be made available to all, including those with limited resources. The OA IJA does not enforce author or subscription fees and will endeavor to obtain alternative sources of income to support this policy for as long as possible.

Editor-in-Chief

Dan Mires

Editorial Board

Rina Chakrabarti	Aqua Research Lab, Dept. of Zoology, University of Delhi, India
Angelo Colorni	National Center for Mariculture, IOLR Eilat, Israel
Daniel Golani	The Hebrew University of Jerusalem Jerusalem, Israel
Hillel Gordin	Kibbutz Yotveta, Arava, Israel
Sheenan Harpaz	Agricultural Research Organization Beit Dagan,
Gideon Hulata	Agricultural Research Organization Beit Dagan,
George Wm. Kissil	National Center for Mariculture, IOLR, Eilat, Israel
Ingrid Lupatsch	Swansea University, Singleton Park, Swansea, UK
Spencer Malecha	Dept. of Human Nutrition, Food & Animal Sciences, CTAHR, University of Hawaii
Constantinos Mylonas	Hellenic Center for Marine Research, Crete, Greece
Amos Tandler	National Center for Mariculture, IOLR Eilat, Israel
Emilio Tibaldi	Udine University Udine, Italy
Jaap van Rijn	Faculty of Agriculture, The Hebrew University of Jerusalem, Israel
Zvi Yaron	Dept. of Zoology, Tel Aviv University, Tel Aviv, Israel

Published under auspices of The Society of Israeli Aquaculture and Marine Biotechnology (SIAMB), University of Hawai'i at Mānoa Library & University of Hawai'i at Mānoa Aquaculture Program in association with AquacultureHub http://www.aquaculturehub.org







ISSN 0792 - 156X

© Israeli Journal of Aquaculture - BAMIGDEH.

PUBLISHER:

Israeli Journal of Aquaculture - BAMIGDEH -Kibbutz Ein Hamifratz, Mobile Post 25210, ISRAEL Phone: + 972 52 3965809 <u>http://siamb.org.il</u>



The *IJA* appears exclusively as a peer-reviewed on-line open-access journal at <u>http://www.siamb.org.il</u>. To read papers free of charge, please register online at <u>registration form</u>. Sale of *IJA* papers is strictly forbidden.



Effects of Dietary Squid Soluble Fractions on Growth Performance and Feed Utilization in Juvenile Snakehead (Ophiocephalus argus) Fed Practical Diets

Biao Yun^{1,2}, Qinghui Ai¹, Hua Xue², Xueqiao Qian^{2*}

¹The Key Laboratory of Mariculture (Education Ministry of China), Ocean University of China, Qingdao, 266003, P.R. China

²Haid Group, Guangdong, China

(Received 10.10.13, Accepted 26.12.13)

Keywords: snakehead; squid soluble fraction; practical diets; growth

Abstract

An 8-week growth trial was conducted to investigate the effects of dietary squid soluble fractions (SSF) supplementation on growth performance and feed utilization of juvenile snakehead (*Ophiocephalus argus*). A fish meal diet (diet 1) with 36% fish meal (FM) was formulated. This diet was used as a control. The second diet (diet 2) was formulated by replacing 22% of the fish meal protein with poultry by-product meal based on amino acid profile of whole body of this fish. The other three diets were supplemented with 0.4% (diet 3), 0.8% (diet 4) and 1.2% (diet 5) SSF, respectively. Diets 1(control) 2, 3, 4, and 5 were isonitrogenous and isolipidic. No significant differences were observed in survival, feed intake (FI), weight gain rate (WGR), feed efficiency rate (FER) and protein productive value (PPV) between fish fed diets 1 and 2 (P>0.05). Furthermore, compared with fish fed the diet without SSF (diet 2), fish fed diets with 0.8 and 1.2% SSF (diet 4 and 5) demonstrated significantly enhanced WGR, FER, and PPV. These results suggest that 0.8% of dietary SSF is optimal for juvenile snakehead growth enhancement.

* Corresponding author: Professor Xue-Qiao Qian; Tel/Fax: +86 20 39990133; email: xueqiaoqian@263.net

Biao et al.

Introduction

Widely farmed snakehead (*Ophiocephalus argus*), is an important commercial carnivorous fish in China and south-east Asian countries (Yu et al., 2012). It is valued for its flesh texture and taste, rapid growth, and extreme tolerance to inferior water quality. Current diets used in snakehead farming are based on high-quality fish meal (FM) as the main protein source. High demands and limited supply have led to high costs for FM. Replacing some FM with animal or plant protein ingredients has become a focal issue (Gatlin et al., 2007; Tacon and Metian, 2008; Gümüş et al., 2013). Terrestrial animal by-products, such as poultry by-product meal (PBM), and meat and bone meal (MBM) are considered to be to be potentially alternative protein sources in aquatic animal feeds due to their high protein content, low carbohydrate content, optimal amino acid profiles, and lack of anti-nutritional factors (Tacon, 1993; Aydin and Gümüş, 2013). These products have also been investigated for a wide range of carnivorous fish species, such as rainbow trout (Bureau et al., 2000), hybrid striped bass (Gaylord and Rawles, 2005), cuneate drum (Wang et al., 2006), Malabar grouper (Wang et al., 2008) and Siberian sturgeon (Zhu et al., 2011; Xue et al., 2012).

Squid soluble fraction is one of the water soluble fractions with low molecular weight proteins, peptides, free amino acids, and nitrogenous compounds (Li et al., 2009; Kousoulaki et al., 2012). Dietary addition of water soluble fractions significantly improved weight gain rate (WGR) in Japanese seabass (Liang et al., 2006), rainbow trout (Aksnes et al., 2006), and Atlantic salmon (Kousoulaki et al., 2009). The small peptides of fish protein hydrolysate, with <100 Da (29.6%) and 1,000-100 Da (18.4%) induced a significant increase in feed consumption in Atlantic salmon and increased fish growth (P<0.05) (Kousoulaki et al., 2a012).

There have been no reports regarding the dietary effect of SSF on growth performance and feed utilization of juvenile snakehead. The objective of the present study is to investigate the effects of dietary SSF supplementation on growth performance and feed utilization of juvenile snakehead fed a practical diet.

Materials and Methods

Feed Ingredients and Diet Formulation. Squid soluble fractions (SSF) was obtained from commercial sources (Haid group Co., Ltd, Guangdong, China). FM, MBM, PBM, and soybean meal (SBM) were used as the primary protein sources (Table 1).

Peptide size (Da)	Squid soluble protein
>70,000	6.00
70,000-50,000	12.80
50,000-20,000	8.40
20,000-10,000	6.90
10,000-5,000	7.10
5,000-1,000	3.80
1,000-100	20.40
<100	34.60

Table 1. Peptide molecular size distribution (% protein) of the squid soluble fractions

Fish oil and soybean lecithin were used as lipid sources. Wheat flour was used as the carbohydrate source. Lysine-HCl and DL-methionine were supplemented to meet the essential amino acid (EAA) requirements of juvenile snakehead. These were based on the whole body amino acid profile (Table 2 and 3). A fish meal diet (diet 1) with 36% FM was formulated and used as the control. The second diet (diet 2) was formulated by replacing 22% of the fish meal protein with poultry by-product meal with an amino acid profile similar to that of the whole body of snakehead. The other three diets were supplemented with 0.4% (diet 3), 0.8% (diet 4) and 1.2% SSF (diet 5), respectively. They were isonitrogenous and isolipidic.

			Diet	no.		
Ingredients	Diet 1	Diet 2	Diet 3	Diet 4	Diet 5	¹ Fishmeal: steam dried fishmeal,
Fish meal ¹	36.0	28.0	27.6	27.2	26.8	(COPENCA Group. Lima, Peru, dry matter basis), with crude protein:
Meat and bone meal ²	4.0	4.0	4.0	4.0	4.0	70.1%, crude lipid: 9.6%;
Poultry by-product meal ³		8.0	8.0	8.0	8.0	² Meat and bone meal composition (dry
Squid soluble fractions			0.4	0.8	1.2	matter basis): crude protein, 70.3%;
Corn gluten meal	4.0	4.0	4.0	4.0	4.0	crude lipid, 13%;
Soybean meal	24.0	24.0	24.0	24.0	24.0	³ Poultry by-product meal composition
Wheat flour	22.0	22.0	22.0	22.0	22.0	(dry matter basis): crude protein, 70 %; crude lipid, 12%;
Fish oil	4.5	4.3	4.3	4.2	4.2	⁴ Squid soluble fractions (dry matter
Soybean lecithin	1.0	1.0	1.0	1.0	1.0	%): crude protein, 72 %; crude lipid,
$Ca(H_2PO_3)_2$	2.0	2.0	2.0	2.0	2.0	14%;
NaCl	0.3	0.3	0.3	0.3	0.3	⁵ Vitamin premix supplied the diet with
Vitamin premix ⁵	0.85	0.85	0.85	0.85	0.85	(mg kg ⁻¹ diet) the following compounds:
Mineral premix ⁶	0.5	0.5	0.5	0.5	0.5	retinyl acetate, 32; vitamin D ₃ , 5; DL-a- tocopherol acetate, 240; vitamin K ₃ , 10;
Ethoxyquin (60%)	0.01	0.01	0.01	0.01	0.01	thiamin, 25; riboflavin (80%), 45;
Lys-HCl		0.40	0.40	0.40	0.40	pyridoxine hydrochloride, 20; vitamin B_{12}
DL-Methionine		0.20	0.20	0.20	0.20	(1%), 10; L-ascorbyl-2-
Rice bran	0.69	0.44	0.44	0.54	0.54	monophosphate-Na (35%), 2000;
Analyzed nutrients compo	sitions (d	ry matter	basis)			calcium Pantothenate, 60; nicotinic acid, 200; inositol, 800; biotin (2%), 60; folic
Dry matter	93.6	93.5	93.7	93.4	93.2	acid, 20; choline chloride (50%), 2500;
Crude protein	47.9	47.9	47.4	47.6	47.4	cellulose, 2473.
Crude lipid	11.5	11.7	11.7	11.7	11.7	⁶ Mineral premix consisted of (mg kg ⁻¹
Ash	11.6	11.3	11.2	11.2	11.1	diet) the following ingredients:
						$FeSO_4 \cdot H_2O, 80; ZnSO_4 \cdot H_2O, 50;$
						CuSO ₄ ·5H ₂ O, 10; MnSO ₄ ·H ₂ O, 45; KI, 60; CoCl ₂ ·6H ₂ O (1%), 50;
						N_{1} , N_{2} , N
						1023003(170), 20, 10304.7120,

Table 2. Formulation & proximate composition of experimental diets (%)

Table 3 Analyzed amino a	cid composition	of the evnerimental	diate (% dry waight)
		or the experimental	

Essential			Diet no.			_
amino acid	Diet 1	Diet 2	Diet 3	Diet 4	Diet 5	47% whole body protein
Thr	1.96	1.89	1.95	1.91	1.91	1.82
Val	2.09	2.09	2.07	2.02	2.03	1.82
Met	1.01	1.04	1.03	1.04	1.04	1.05
Ile	1.92	1.84	1.87	1.82	1.81	1.61
Leu	3.45	3.35	3.43	3.34	3.30	2.95
Phe	2.08	2.07	2.10	2.02	2.02	1.67
Lys	3.21	3.30	3.37	3.28	3.33	3.23
His	1.33	1.25	1.27	1.23	1.25	0.87
Arg	2.73	2.69	2.74	2.66	2.71	2.59

Ingredients were ground into fine powder and sieved through 246 μ m mesh. All the ingredients were then thoroughly mixed with menhaden fish oil, and water was added to produce stiff dough. The dough was then pelleted using an experimental feed mill (F-26 (II), South China University of Technology, China) and dried for about 12h in a ventilated oven at 45°C, and then kept frozen at -20°C.

Fish, Experimental Conditions and Samples Collection. Juvenile snakehead was obtained from the Haid fish farm (Panyu, Guangdong, China). Fish were acclimated to the system and fed diet 1 (control) for 2 weeks before the commencement of the trials. Juvenile snakehead (initial body weight: 22.3 ± 0.1 g) were randomly distributed into 15 flat bottomed tanks, filled with 3200 I freshwater which was continuously pumped from a pond adjacent to the experiment station, and sieved through separate sand filters into each tank at approximately 1.5 I/min. Three replicate tanks were randomly assigned to each diet group and 120 fish were bulk weighed and stocked in each tank. During the 8 week feeding period, fish were fed the experimental diets to apparent satiation twice

1200; calcium propionate, 1000;

zoelite, 2485.

daily at 0700 and 1700 h respectively. Any uneaten feed was collected 1 h after each meal, dried to constant weight at 70°C and reweighed. Leaching loss in the uneaten diet was estimated by leaving five samples of each diet in tanks without fish for 1 h, recovering, drying, and reweighing.

Before the experiment, 10 fish from the same population were randomly selected for determination of initial whole-body proximate composition. At the end of the experiment, 6 fish weighing approximately the same as fish in the experimental groups were sampled and stored frozen at -20° C for whole body composition analysis. During the 8-week feeding period, water temperature ranged from 26 to 30° C, pH 7.5-8.0, ammonia nitrogen was lower than 0.1 mg/l, nitrite was lower than 0.1 mg/l, and dissolved oxygen was higher than 6.0 mg/l.

Chemical analyses. Fish samples, ingredients, and experimental diets were analyzed for dry matter, crude protein, crude lipid, and ash. (AOAC, 1995).

Dry matter was analyzed by drying the samples to constant weight at 105°C. Crude protein was determined using the Kjeldahl method and estimated by multiplying nitrogen by 6.25. Crude lipid was measured after diethyl ether extraction using Soxhlet method. Ash was examined by combustion in a muffle furnace at 550°C for 16 h. Duplicate analyses were conducted for each sample.

The SSF of the experimental diets was extracted with boiling water, the extract was then filtered through Black ribbon filter paper and the crude protein content in the water phase was determined using the Kjeldahl method. Size distribution of peptides was analyzed by HPLC size exclusion chromatography using a TSK G2000 column and detection at 220 nm according to Aksnes and Asbjørnsen (2003).

Calculations and Statistical Methods. Growth parameters were calculated as follows: Weight gain rate (WGR) (%) = $100 \times [(final body weight - initial body weight) / initial$

body weight]

Feed intake (FI) $(\%/d) = 100 \times \text{total}$ amount of the feed consumed (g) / [(initial body weight + final body weight) / 2] / days

Feed efficiency rate (FER) = wet weight gain (g) / total amount of the feed consumed (g) Protein productive value (PPV) (%) = $100 \times \text{ body wet protein gain (g)/ protein intake (g)}$ Survival rate (SR) (%) = $100 \times \text{(final fish number / initial fish number)}$.

The Software SPSS, 11.5 was used for all statistical evaluations. All data were subjected to one-way analysis of variance (ANOVA) followed by Tukey's test. Differences were regarded as significant when P<0.05. Data are expressed as means ± standard error.

Results

All experimental diets were well accepted by the fish. No pathological signs were observed during the 8-week feeding period.

Squid Soluble Fractions Composition. The peptide size distribution of SSFs are given in Table 1.

Survival Rates and Growth Performance. Survival rate was higher than 95% in all treatments and no significant differences (P>0.05) were found between the treatments (Table 4).

Table 4 Growth performance and survival rate of snakehead fed the experimental diets (Means \pm SE, n = 3)*

			Diet no.		
	Diet 1	Diet 2	Diet 3	Diet 4	Diet 5
IBW ¹	22.3±0.1	22.3±0.1	22.3±0.1	22.3±0.1	22.3±0.1
FBW ²	115.6±3.1ª	117.8±2.6ª	120.0±1.9 ^{ab}	124.7±3.1 ^b	125±1.0 ^b
WGR ³	419.5±14.1ª	429.3±11.7ª	439.3±8.5 ^{ab}	460.3±13.7 ^b	461.8±4.5 ^b
FI^4	2.39±0.02	2.38±0.02	2.32±0.08	2.19±0.06	2.22±0.02
FER⁵	0.94±0.02 ^b	0.95 ± 0.02^{b}	0.99 ± 0.03^{ab}	1.06±0.03ª	1.05 ± 0.01^{a}
SR ⁶	98.3±0.8	99.2±0.8	95.0±2.5	99.2±0.8	97.5±1.4

¹ IBW: initial body weight; ² FBW: final body weight

³ Weight gain rate (WGR) (%) = $100 \times [(FBW - IBW) / IBW]$

⁴ Feed intake (FI) (%/d) = $100 \times \text{total amount of the feed consumed / [(IBW + FBW) / 2] /days$

⁵ Feed efficiency rate (FER) = wet weight gain (g) / total amount of the feed consumed (g)

⁶ Survival rate (%) = $100 \times$ (final fish number / initial fish number)

*Values in the same row with different superscripts are significantly different (P<0.05)

The feed intake (FI) of fish ranged from 2.19%/day to 2.39%/day (P>0.05), without any significant differences between dietary treatments. There were also no significant differences in weight gain rate (WGR), feed efficiency rate (FER) and protein productive value (PPV) between diet 1 and diet 2 (P>0.05). Furthermore, compared with fish fed the diet without SSF (diet 2), and those fed diets with higher SSF (diets 4 and 5) had significantly enhanced WGR, FER, and PPV (P<0.05) Fig 1.

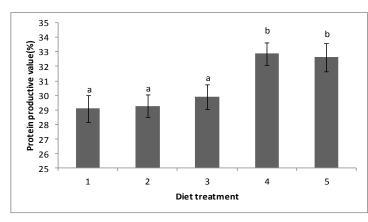


Fig 1. Protein productive value (PPV) of snakehead fed the experimental diets. Data of the five experimental treatments are presented as mean \pm S.E. (n=3). Different letters above the bars denote significant differences among treatments at the *P*<0.05 level.

Body Composition. No significant differences (P>0.05) were detected between dietary treatments with respect to wholebody moisture (73.8-75.4%), crude protein (14.6-15.0%), crude lipid (7.7-8.2%), and ash

(3.5-3.8%) contents of fish (Table 5).

Table 5. Proximate composition in whole body of snakehead fed the experimental diets (% diet in wet basis; means \pm SE, n = 3)*

	Diet no.					
	Diet 1	Diet 2	Diet 3	Diet 4	Diet 5	
Moisture	75.0±0.4	74.9±0.8	73.8±1.0	74.6±1.1	75.4±0.8	
Crude protein	14.7±0.2	14.7±0.2	15.0 ± 0.9	14.7±0.6	14.6±0.6	
Crude lipid	7.7±0.5	7.9±0.6	8.2±0.8	7.9±0.4	7.7±0.2	
Ash	3.8±0.2	3.5±0.4	3.6±0.4	3.8±0.2	3.8±0.2	_

*Values in the same row with different superscripts are significantly different (P<0.05).

Discussion

Limited supply, increasing demand, and high prices of FM are challenges to the sustainable development of the fish culture industry, notably carnivorous fish which are frequently fed feeds with high FM levels (Wang et al., 2006; Glencross et al., 2007). Cost effectiveness of feed could be improved by replacing FM with more economical protein sources. Numerous studies have focused on assessing the potential to reduce FM level in fish feeds. The present study shows that there was no significant difference in the growth of snakehead with an initial body weight of 22.3 g when fish meal protein was replaced by 22% poultry by-product meal. An optimal EAA profile is a prerequisite for fish growth and nitrogen retention (Luo et al., 2006; Peres and Oliva-Teles, 2009). The content of the EAA, especially the Lysine (Lys), Methionine (Met), and Threonine (Thr) content, is generally the limiting amino acid content in economical alternative protein sources. A deficiency in one EAA will lead to poor utilization of the provided dietary protein (Wilson, 2002). In the present study, Lysine-HCl and DL-methionine (Crystalline amino acids) were supplemented so that the EAA requirements of juvenile snakehead were met based on the whole-body amino acid profiles. The dietary FM levels were reduced to 28% by incorporating poultry by-product meal based on whole-body amino acids profile without negative effects on growth performance and feed utilization.

In the present study, the growth rate of fish fed diets with 0.8% or 1.2% SSF was significantly better (P<0.05) compared with that of fish fed the diet without SSF (diet 2). Similar results have been observed in Atlantic salmon (Espe et al., 1999), rainbow trout (Aksnes et al., 2006) and Japanese seabass (Liang et al., 2006) fed the small peptide fraction of fish protein hydrolysate. In this study, the inclusion of SSF enhanced the effect of bioactive nitrogenous compounds such as small peptides present in high amounts in the soluble part of marine raw materials (Table 1). Significant effects of fish hydrolysate

Biao et al.

fraction have been observed on the spatial expression of peptide transporter in the digestive tract of Atlantic cod (Bakke et al. 2010). This may also suggest that the growth promoting effect of SSF may be partly attributed to the modification in the nutrient absorption patterns in fish. However, a diet with excessive SSF may retard growth in larvae or juvenile fish, such as in larvae of sea bass (Cahu et al., 1999), turbot juveniles (Oliva-Teles et al., 1999), larvae of cobia (Han et al., 2010), and larvae of tongue sole (Liu et al., 2010). Some of these detrimental effects may be linked to a fast flow of short peptide and free amino acids through the gut wall, a flow that the larvae cannot handle. As a result, most of these metabolites could be flushed out of the digestive system (Espe and Njaa, 1991; Kolkovski and Tandler, 2000).

In the present study, no significant differences were detected between dietary treatments with respect to whole-body moisture, crude protein, crude lipid, and ash content of fish (P>0.05). These results are in agreement with previous studies on Atlantic salmon (Kousoulaki et al., 2009, 2012) and Japanese flounder (Zheng et al., 2011).

In conclusion, results of the present study show that: dietary FM levels can be reduced to 28% by incorporating poultry by-product without negative effects on growth performance and feed utilization; and 0.8% of dietary SSF seems optimal for significantly better growth rate and feed utilization of juvenile snakehead.

Acknowledgements

This research was supported by the Guangdong Marine economy development demonstration area special project (GD2012—B01-010).

References

Aksnes A. and B. Asbjørnsen, 2003. Method for the definition of peptide size in fish hydrolysate, ensilage and gelatine. Fiskeriforskning Report K-289 (in Norwegian).

Aksnes A., Hope B., Jönsson E., Björnsson B.T. and S. Albrektsen, 2006. Sizefractionated fish hydrolysate as feed ingredient for rainbow trout (*Oncorhynchus mykiss*) fed high plant protein diets. I: growth, growth regulation and feed utilization. *Aquaculture*, 261:305-317.

Aydin B. and E. Gümüş, 2013. Replacement of Fish Meal by Poultry By-product Meal with Lysine, Methionine and Threonine Supplementation to Practical Diets for Nile Tilapia Fry (*Oreochromis niloticus*), *Isr. J. Aquacult. - Bamidgeh*, IJA_65.2013.885, 7 pages.

Association of Official Analytical Chemist (AOAC), (1995) Official Methods of Analysis of AOAC International, 16th edn. AOAC Inc., Arlington, VA, USA.

Bakke S., Jordal A.E.J., Gómez-Requeni P., Verri T., Kousoulaki K., Aksnes A. and I. Rønnestad, 2010. Dietary protein hydrolysates and free amino acids affect the spatial expression of peptide transporter PepT1 in the digestive tract of Atlantic cod (*Gadus morhua*). *Comp. Biochem. Physiol*.156:48-55.

Bureau D.P., Harris A.M., Bevan D.J., Simmons L.A., Azevedo P.A. and C.Y. Cho, 2000. Feather meals and meat and bone meals from different origins as protein sources in rainbow trout (*Oncorhynchus mykiss*) diets. *Aquaculture*, 181:281-291.

Cahu C, Zambonino Infante J.L. and P. Quazuguel, 1999. Protein hydrolysate vs. fish meal in compound diets for 10-day old sea bass *Dicentrarchus labrax* larvae. *Aquaculture*, 171:109-119.

Espe M. and L.R. Njaa, 1991. Growth and chemical composition in Atlantic salmon (*Salmo salar*) given a fish meal diet or corresponding free amino acid diet. *Fisk. Dir. Skr. Ser. Ernaering*, 4:103-110.

Espe M., Sveier H., Hogoy I. and E. Lied, 1999. Nutrient absorption and growth of Atlantic salmon (*Salmo salar* L.) fed fish protein concentrate. *Aquaculture*, 174:119-137.

Gatlin III D.M., Barrows F.T. and P. Brown, 2007. Expanding the utilization of sustainable plant products in aquafeeds a review. *Aquacult. Res.*, 38:551-579.

Gaylord T.G. and S.D. Rawles, 2005. The modification of poultry by-product meal for use in hybrid striped bass diets. *J. World Aquacult. Soc.*, 36:365-376.

Glencross B.D., Booth M. and G.L. Allan, 2007. A feed is only as good as its ingredients - a review of ingredient evaluation strategies for aquaculture feeds. *Aquacult. Nutr.*, 13:17-34.

Gümüş E., Erdoğan F. and B. Aydın, 2013. Evaluation of Skate Meal as a Replacement of Fishmeal in Diets for Nile Tilapia Fry (*Oreochromis niloticus*), <u>Isr. J. Aquacult.</u> - <u>Bamidgeh</u>, IJA_65.2013.922, 9 pages.

Han T., Wang J.T., Wang Y., Tian L.X. and Y.J. Liu, 2010. Effect of difference fish protein hydrolysate (FPH) level of dietary supplements on growth and body composition of larvae of cobia (*Rachycentron canadum*). *Acta Hydrobiologica sinica*, 34:94-100.

Kolkovski S. and A. Tandler, 2000. The use of squid protein hydrolysate as a protein source in microdiets forgilthead seabream *Sparus aurata* larvae. *Aquacult. Nutr.*, 6:11-15.

Kousoulaki K., Albrektsen S., Langmyhr E., Olsen H.J., Campbell P. and A. Aksnes, 2009. The water soluble fraction in fish meal (stickwater) stimulates growth in Atlantic salmon (*Salmo salar* L.) given high plant protein diets. *Aquaculture*, 289:74-83.

Kousoulaki K., Olsen H.J., Albrektsen S., Langmyhr E., Mjøs S.A., Campbell P. and A. Aksnes, 2012. High growth rates in Atlantic salmon (*Salmo salar* L.) fed 7.5% fish meal in the diet. Micro-, ultra- and nano-filtration of stickwater and effects of different fractions and compounds on pellet quality and fish performance. *Aquaculture*, 338-341:134-146.

Li P., Mai K., Trushenski J. and G. Wu, 2009. New developments in fish amino acid nutrition: towards functional and environmentally oriented aquafeeds. *Amino Acids*, 37:43-53.

Liang M., Wang J., Chang Q. and K. Mai., 2006. Short communication. Effects of different levels of fish protein hydrolysate in the diet on the nonspecific immunity of Japanese sea bass, *Lateolabrax japonicus* (Cuvieret Valenciennes, 1828). *Aquacult. Res.*, 37:102-106.

Liu X.D., Liang M.Q., Zhang L.M., Wang J.Y., Chang Q., Wang J.L. and K. Yannis, 2010. Effect of fish protein hydrolysate levels on growth performance, biological and physiological parameters in tongue sole (*Cynoglossus semiluevis* Gunther, 1873) post-larvae. *Acta Hydrobiologica sinica*, 2:242-249.

Luo L., Xue M., Wu X.F., Cai X.F., Cao H.N. and Y.M. Liang, 2006. Partially or totally replacement of fish meal by solvent-extracted cottonseed meal in diets for juvenile rainbow trout (*Oncorhynchus mykiss*). *Aquacult. Nutr.*, 12:418-424.

Oliva-Teles A., Cerqueira A. L. and P. Gonc alves, 1999. The utilization of diets containing high levels of fish protein hydrolysate by turbot (*Scophthalamus maximus*) juveniles. *Aquaculture*, 179:195-201.

Peres H. and A. Oliva-Teles, 2009. The optimum dietary essential amino acid profile for gilthead seabream (*Sparus aurata*) juveniles. *Aquaculture*, 296:81-86.

Tacon A.G.J., 1993. Feed ingredients for warm water fish, fish meal and other processed feedstuffs. FAO Fisheries Circular, 856, 64p.

Tacon A.G.J. and M. Metian, 2008. Global overview on the use of fish meal and fish oil in industrially compounded aquafeeds: trends and future prospects. *Aquaculture*, 285:146-158.

Wang Y., Kong L.J., Cui Z.H., Li K. and D.P. Bureau, 2006. Effect of replacing fish meal with soybean meal on growth, feed utilization and carcass composition of cuneate drum (*Nibea miichthioides*). *Aquaculture*, 261:1307-1313.

Wang Y., Li K., Han H., Zheng Z.X. and D.P. Bureau, 2008. Potential of using a blend of rendered animal protein ingredients to replace fishmeal in practical diets for malabar grouper (*Epinephelus malabricus*). *Aquaculture*, 281:113-117.

Wilson R. P., 2002. Amino acids and Protein. In: *Fish Nutrition* (Halver, J.E. and Hardy, R.W. eds), 3rd edn, pp. 309-365. Elsevier, Amsterdam, the Netherlands.

Xue M., Yun B., Wang J., Sheng H.J., Zheng Y.H., Wu X.F., Qin Y.C. and P. Li, 2012. Performance, body compositions and metabolism of nitrogen and phosphorus in Siberian sturgeon, *Acipenser baerii* Brandt, as affected by dietary animal protein blend replacing fishmeal and protein levels. *Aquacult. Nutr.*, 18:493-501.

Yu H.R., Zhang Q., Cao H., Wang X.Z., Huang G.Q., Zhang B.R., Fan J.J., Liu S.W., Li W.Z. and Y. Cui, 2012. Apparent digestibility coefficients of selected feed ingredients for juvenile snakehead, *Ophiocephalus argus. Aquacult. Nutr.*, 19:139-147.

Zheng K.K., Liang M.Q., Yao H.B., Chang Q, and J.L. Wang, 2011. Inclusion of sizefractionated fish hydrolysate in high plant protein diets for Japanese flounder, Paralichthys olivaceus. ACTA *Hydrobiol. Sini.*, 35:829-834.

Zhu H., Gong G., Wang J., Wu X.F., Xue M., Niu C.J., Guo L.Y. and Y. Yu, 2011. Replacement of fish meal with blend of rendered animal protein in diets for Siberian sturgeon (*Acipenser baerii* Brandt), results in performance equal to fish meal fed fish. *Aquacult. Nutr.*, 17:e389-e395.