

Relaxation of Selective Constraint on the Ultra-Large Mitochondrial Genomes of Arcidae (Mollusca: Bivalvia)

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Abstract The mitochondrial genomes (mitogenomes) are purportedly under selection for smaller size to improve their replication and translation efficiency. However, the mitogenomes of Arcidae species are larger than those of other bivalves, and are among the largest metazoan mitogenomes reported to date. In order to explore the differences of base composition and selective constraints between the large and small mitogenomes, we compared the mitogenomes of 9 large arcid mitogenomes and 77 small bivalves mitogenomes. Base composition analyses indicated that Arcidae mitogenomes have significantly greater GC skews in both their whole genomes and coding sequences. This result suggests that the replication of the large mitogenomes in Arcidae may be slower than those in other bivalves, exposing the parental strand to deamination for a longer time. Selection pressure analyses showed that the mitochondrial protein-coding genes of Arcidae species have significantly higher Ka/Ks ratios than other bivalves, suggesting that they have accumulated more nonsynonymous nucleotide substitutions. Seven protein-coding genes (*atp6*, *cox1-3*, *nad1*, *nad4* and *nad5*) show significant difference for Ka/Ks ratios between the Arcidae and non-Arcidae groups. However, these divergences are not observed in the nuclear gene within histone H3. From these observations, we concluded that the large mitogenomes of Arcidae species experienced more relaxed selective constraints. As some Arcidae species are more tolerant to hypoxia that can lead to low metabolic rate, the relaxed selective constraints of mitogenomes may be energy-related. This study provides new insights into the evolution of Arcidae mitogenomes.

Key words Arcidae; mitochondrial genome; genome size; relaxed selective constraint

1 Introduction

The mitochondrial genomes (mitogenomes) of most bilaterian animals include a standard set of 13 protein-coding genes (PCGs), 2 ribosomal RNA (rRNA) genes, 22 transfer RNA (tRNA) genes, and an A+T-rich region (Boore, 1999). Although there are exceptions, most mitogenomes size are from 14 to 17kb. Typically, few intergenic nucleotides exist except for a single large non-coding region, which were thought to contain elements that control the initiation of replication and transcription of the mitogenome (Boore, 1999; Lavrov, 2007). This consistency in gene content across distantly related lineages, as well as the lack of intergenic spacers, suggests that the mitogenome is under selection for compact size (Rand and Harrison, 1986). Compared with nuclear genome, the mitogenome has several advantages including conserved gene content, maternal inheritance, lack of extensive recombination, and relatively high nucleotide substitution rates (Boore, 1999; Curole and

Kocher, 1999; Gissi *et al.*, 2008). These advantages make it a good model for the studies of evolutionary genomics (Saccone *et al.*, 1999; Gissi *et al.*, 2008; Cameron, 2014).

Molluscs, especially bivalves, usually display an extraordinary amount of variation in mitogenome structure and size, even with differences in the closely related species (Gissi *et al.*, 2008; Simison and Boore, 2008). The size of the bivalve mitogenomes are highly variable, ranging from 14622 in *Lanternula elliptica* (Park and Ahn, 2015) to 46985 bp in *Scapharca broughtonii* (Liu *et al.*, 2013) in length. Several bivalve species have showed large sizes of mitogenomes (greater than 20kb). The mitogenome size of the deep sea scallop *Placopecten magellanicus* is up to 40725 bp (Smith and Snyder, 2007) and the mitogenome of Zhikong scallop *Chlamys farreri* is 21695 bp (Xu *et al.*, 2011). The mitogenome size of Manila clam *Venerupis philippinarum* is 22676 bp in female type and 21441 bp in male type (Passamonti and Scali, 2001). The mitogenome of *Bryopa lata*, Clavagellidae, is at least 31969 bp long (Williams *et al.*, 2017). The size of Arcidae mitogenomes are more unusual, of which the largest mitogenome size is 46985 bp (*Scapharca broughtonii*) (Liu *et al.*, 2013). The

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bivalve species have a sedentary lifestyle, with low metabolic rate (Sun *et al.*, 2017). The previous researches proposed that the large mitogenome of bivalves perhaps experience weak purifying selection, which may be correlated with their low metabolic rates (Strotz *et al.*, 2018; Kong *et al.*, 2020).

Ark shells are among the oldest bivalve lineages, reaching back to the lower Ordovician which is about 450 Myr (Morton *et al.*, 1998). The species of Arcidae are globally distributed, predominantly in the tropical shallow waters and warm temperate seas, containing approximately 260 species and 31 genera (Oliver and Holmes, 2006). Mitogenomes of ark shell species are among the largest metazoan mitogenomes reported to date, ranging from 18 to 56 kb in length (Kong *et al.*, 2020). It has been argued that animal mitogenomes are characterized by a tightly packed collection of conserved genes and other functional elements, accompanying with drastic mitogenome size reduction in evolutionary history of animal (Burger *et al.*, 2003; Schneider and Ebert, 2004; Signorovitch *et al.*, 2007). Arcidae presented a challenge to the point of selection favoring compact genomes by virtue of large mitogenome size. In our previous studies, we found the mitogenome size is positively correlated with the combined length of *cox1*, *cox2* and *cox3*, the length of *Cytb*, and the combined length of *rrnS* and *rrnL* (Sun *et al.*, 2016). Researchers have found the inverted repeat sequences might facilitate the mitogenome expansions (Kong *et al.*, 2020). It has been believed that metabolic rates influence the selective constraints acting on the mitogenome, and purify selection act on small genome size (Rand, 1993). However, the natural selection act on the large mitogenome of Arcidae remains unexplored.

The large size of Arcidae mitogenomes lead to several questions. Firstly, whether the nucleotide compositions of large Arcidae mitochondrial DNA (mtDNA) are different from those of other bivalve species with small ones? Secondly, what's the difference in substitution rate of mtDNA between Arcidae and other bivalve species? Thirdly, do all the mitochondrial genes of Arcidae experience the same selection pressure? In order to address these questions, we conducted a comparative genomic analysis and test the roles of the evolutionary constraints on the mtDNA of Arcidae to provide a complete view of molecular evolution in the mtDNA.

2 Materials and Methods

2.1 Source of Data

The mtDNA sequences (Table 1) and nuclear gene (histone H3) (Table 2) of bivalves were downloaded from GenBank. All the mitochondrial protein-coding genes were extracted from each mitogenome.

2.2 Base Composition

AT and GC skew were calculated according to the formula defined by Perna and Kocher (1995), AT skew = $(A - T)/(A + T)$ and GC skew = $(G - C)/(G + C)$, which provides an index of compositional asymmetry between strands. Skews were calculated for all sites, and also for fourfold degenerate sites, which are expected to be less constrained (Reyes *et al.*, 1998). We then compared these measures of nucleotide skew between Arcoidae and non-Arcoidae taxa. All statistical analyses were performed with IBM SPSS Statistics, release 19.0.0.1.

Table 1 The mtDNA sequences of bivalves downloaded from GenBank

Species	Accession no.	Ka/Ks	Ka	Ks	<i>atp6</i>	<i>cox-1</i>	<i>cox-2</i>	<i>cox-3</i>	<i>cytb</i>	<i>nd1</i>	<i>nd2</i>	<i>nd3</i>	<i>nd4</i>	<i>nd4l</i>	<i>nd5</i>	<i>nd6</i>
<i>Tegillarca granosa</i>	KJ607173	0.0552	0.0492	0.8918	0.0284	0.0447	0.1107	0.0247	0.0255	0.0517	0.0967	0.0196	0.0526	0.0418	0.0895	0.0641
<i>Scapharca kagoshimensis</i>	KF750628	0.1188	0.0741	0.6240	0.0203	0.0478	–	0.1438	0.2349	0.0874	0.9466	0.2834	0.0357	0.0414	0.2071	0.1105
<i>Scapharca broughtonii</i>	AB729113	0.0901	0.0147	0.1635	0.6263	0.0883	0.0418	–	–	0.0303	0.0745	–	–	0.0360	0.0918	–
<i>Anadara vellicata</i>	KP954700	0.0558	0.0654	1.1734	0.0909	0.0125	0.0738	0.0351	0.0362	0.1339	0.9985	0.0479	0.0710	0.0372	0.0714	0.1068
<i>Trisidos kiyoni</i>	KU975161	0.0434	0.2155	4.9697	0.1043	0.0265	0.0330	0.1235	0.0262	0.0664	0.1905	0.1165	0.0882	0.0341	0.0353	0.0794
<i>Potiarca pilula</i>	KU975162	0.0681	0.1364	2.0028	0.0466	0.0352	0.1266	0.0497	0.0507	0.0445	0.0048	0.0448	0.0862	0.3316	0.0824	0.0692
<i>Arca navicularis</i>	MG641752	0.0531	0.2711	5.1060	0.0828	0.0320	0.0642	0.1479	0.1805	0.0839	0.0456	0.0690	0.0838	0.1316	0.0613	0.1800
<i>Barbatia virescens</i>	MF374794	0.0845	0.0865	1.0236	0.0745	0.0325	0.1254	0.0754	0.0512	0.0521	0.0853	–	0.0853	0.2313	0.0863	0.0741
<i>Anadara consociata</i>	MG641753	0.0654	0.0717	1.0961	0.0887	0.0230	0.1783	0.0506	0.0200	0.0353	0.1255	0.0528	0.0381	0.0191	0.0735	0.0870
<i>Crassostrea hongkongensis</i>	FJ841963	0.0433	0.0156	0.3608	0.0190	0.0067	0.0001	0.0236	0.0468	0.0453	0.0890	0.4552	0.0638	0.0187	0.0846	0.0561
<i>Crassostrea iredalei</i>	FJ841967	0.0571	0.0510	0.8927	0.0338	0.0091	0.0121	0.0872	0.0868	0.0743	0.1286	0.0664	0.0835	0.0140	0.0656	0.0651
<i>Crassostrea sikamea</i>	FJ841966	0.0513	0.0158	0.3073	0.0394	0.0001	0.0001	0.0194	0.0537	0.0266	0.0546	0.1069	0.0661	0.0001	0.1571	0.0583
<i>Crassostrea angulata</i>	FJ841965	0.0481	0.0027	0.0569	0.0001	0.0001	0.0001	0.1289	0.0317	0.0598	0.2178	0.3846	0.0432	0.2252	0.0913	0.0677
<i>Crassostrea ariakensis</i>	FJ841964	0.0851	0.0439	0.5154	0.0036	0.0122	0.0038	0.0722	0.1402	0.0550	0.2060	0.1997	0.0935	0.1377	0.1070	0.3430
<i>Crassostrea virginica</i>	AY905542	0.0161	0.0669	4.1576	0.0209	0.0116	0.0155	0.0465	0.0271	0.0201	0.0322	0.0334	0.0130	0.0937	0.0438	0.0128
<i>Crassostrea gigas</i>	KJ855241	0.1020	0.0063	0.0615	0.5212	0.0125	0.0001	0.0215	0.0401	0.2728	0.1779	0.0503	0.1310	0.0001	0.2280	0.3055
<i>Crassostrea gasar</i>	KR856227	0.0124	0.0578	4.6675	0.0208	0.0099	0.0026	0.0077	0.0173	0.0145	0.0283	0.0783	0.0144	0.0285	0.0528	0.0503
<i>Crassostrea nippona</i>	HM015198	0.0621	0.0383	0.6162	0.0289	0.0185	0.0427	0.1081	0.0546	0.0350	0.1143	0.0565	0.0826	0.0508	0.1148	0.0694
<i>Ostrea lurida</i>	KC768038	0.0506	0.0301	0.5939	0.0053	0.0049	0.0103	0.0864	0.0530	0.0556	0.0414	0.0883	–	–	0.0586	0.1849
<i>Ostrea edulis</i>	JF274008	0.0362	0.0429	1.1847	0.0222	0.0468	0.0260	0.0512	0.0793	0.0138	–	0.0215	0.0303	0.0271	0.0263	0.0665
<i>Saccostrea mordax</i>	FJ841968	0.0230	0.0324	1.4093	0.0293	0.0132	0.0056	0.0374	0.0287	0.0179	0.0659	0.0247	0.0263	–	0.0229	0.0399
<i>Saccostrea cucullata</i>	KP967577	0.0245	0.0263	1.0749	0.0158	0.0008	0.0191	0.0191	0.0234	0.0181	0.0515	0.0013	0.0212	0.0071	0.0562	0.1323
<i>Mimachlamys nobilis</i>	FJ415225	0.0400	0.0378	0.9460	0.0708	0.0134	0.0648	0.0378	0.0397	0.0167	0.0419	0.1007	0.0272	0.0065	0.0644	0.0567
<i>Mimachlamys senatoria</i>	KF214684	0.0261	0.0227	0.8678	0.0273	0.0129	0.0280	0.0242	0.0192	0.0126	0.0511	0.7871	0.0197	0.0203	0.0589	0.0226
<i>Placopecten magellanicus</i>	DQ088274	0.0545	0.1395	2.5600	0.1003	0.0310	0.0557	0.0453	0.0539	0.0315	0.0820	0.0780	0.0654	0.1249	0.1613	0.1204

(to be continued)

Table 2 The nuclear gene (histone H3) of bivalves downloaded from GenBank

Species	Accession no.	Ka/Ks	Species	Accession no.	Ka/Ks
<i>Crassostrea ariakensis</i>	KM460854	0.0001	<i>Scapharca subcrenata</i>	JN974603	0.0001
<i>Crassostrea gigas</i>	KM460855	0.0001	<i>Scapharca broughtonii</i>	JN974601	0.0001
<i>Crassostrea virginica</i>	KM460872	0.0001	<i>Scapharca inaequivalvis</i>	JN974599	0.0001
<i>Crassostrea nippona</i>	KM460859	0.0001	<i>Scapharca cornea</i>	JN974597	0.0001
<i>Ostrea circumpecta</i>	KM460864	0.0298	<i>Scapharca gubernaculum</i>	JN974594	0.0001
<i>Ostrea denselamellosa</i>	KM460862	0.0771	<i>Anadara antiquata</i>	JN974592	0.0001
<i>Saccostrea kegaki</i>	KM460867	0.0001	<i>Anadara crebricostata</i>	JN974591	0.0001
<i>Pinctada martensi</i>	JN974634	0.0001	<i>Anadara vellicata</i>	JN974589	0.0001
<i>Pinctada margaritifera</i>	KU763362	0.0001	<i>Mytilisepta virgata</i>	MK642879	0.0001
<i>Argopecten purpuratus</i>	EU379525	0.0001	<i>Modiolus modiolus</i>	HF545159	0.0001
<i>Pseudamussium peslutrae</i>	EU379529	0.0001	<i>Bathymodiolus manusensis</i>	HF545158	0.0001
<i>Argopecten gibbus</i>	EU379497	0.0001	<i>Bathymodiolus boomerang</i>	HF545135	0.0001
<i>Mimachlamys asperrima</i>	HM540086	0.0001	<i>Bathymodiolus brevior</i>	HF545132	0.0001
<i>Arca</i> sp.	JN974630	0.0001	<i>Mytilus edulis</i>	AY377769	0.0001
<i>Barbatia virescens</i>	KT757878	0.0001	<i>Trachycardium egmontianum</i>	KR422792	0.0001
<i>Barbatia fusca</i>	JN974626	0.0001	<i>Vasticardium angulatum</i>	KR422797	0.0001
<i>Trisidos kiyonoi</i>	JN974623	0.0001	<i>Vasticardium assimile</i>	KR422799	0.0001
<i>Arca navicularis</i>	JN974618	0.0001	<i>Lyrocardium lyratum</i>	KR422775	0.0001
<i>Barbatia lima</i>	JN974613	0.0001	<i>Laevicardium serratum</i>	KR422774	0.0001
<i>Scapharca globosa</i>	JN974584	0.0001	<i>Laevicardium pictum</i>	KR422772	0.1361
<i>Arca ventricosa</i>	AF416858	0.0001	<i>Keenocardium blandum</i>	KR422767	0.0001
<i>Barbatia lacerata</i>	JN974611	0.0001	<i>Fulvia undatopicta</i>	KR422766	0.0001
<i>Anadara pilula</i>	JN974608	0.0001	<i>Fulvia nienkeae</i>	KR422763	0.0001
<i>Tegillarca granosa</i>	JN974607	0.0001	<i>Fulvia mutica</i>	KR422761	0.0001
<i>Tegillarca nodifera</i>	JN974605	0.0001			

2.3 Estimation of Nonsynonymous/Synonymous Substitutions Ratios (Ka/Ks)

The maximum-likelihood phylogenetic relationships were reconstructed based on nucleotide sequences of twelve protein-coding genes using RAxML v.7.0.4 (Stamatakis, 2006). The twelve-partitioned nucleotide sequences were aligned with ClustalX (Thompson *et al.*, 1997). The ratios of nonsynonymous to synonymous substitutions (*Ka/Ks*) were estimated for each branch using CodeML implemented in the PAML package (Yang, 2007). Model 1 was used, which allows a free *Ka/Ks* ratio. Only *Ka/Ks* and *Ks* values of the external branches were selected in the following analyses, *i.e.*, deleterious mutations (*Ka/Ks*) between modern species and their most recent ancestors. The statistical analyses were performed with IBM SPSS Statistics, release 19.0.0.1.

3 Results and Discussion

3.1 Relationship Between Genome Size and Nucleotide Composition in Arcidae

Arcidae species possess larger mitogenomes than that found in typical animals, challenging the conventional hypothesis that a compact mitogenome is a common feature among all animals. The increased size of Arcidae mitogenome is due to the presence of long noncoding regions. Genomic coverage by mitochondrial noncoding regions can reach up to 71% (33046bp) for *S. kagoshimensis* (Sun *et al.*, 2014). Larger size molecules are usually considered to be at a selective disadvantage simply because they take

longer time to replicate, leaving fewer copies to be transmitted (Boyce *et al.*, 1989). On the contrary, smaller sized mtDNA molecules are with replicative or selective advantage (Boyce *et al.*, 1989).

In order to explore if the large Arcidae mtDNA can affect replication mechanics, we compared the nucleotide skew of whole genome sequences (P_{mtDNA}), the protein-coding genes at all codon positions (P_{123}), and the four-fold codon positions ($P_{4\text{FD}}$) between Arcidae species and the other bivalve species (Table 3). AT skews for the P_{mtDNA} are the same between Arcidae and non-Arcidae groups (Mann-Whitney U-test, $P=0.904$, Fig. 1A); however, the GC skews of Arcidae species are significantly greater than that of the non-Arcidae group ($P=0.003$, Fig. 1B). The GC skews of P_{123} in Arcidae species are also significantly greater than those of non-Arcidae group ($P=0.001$, Fig. 1C), while AT skews do not differ ($P=0.076$, Fig. 1D). This pattern is similar when analysis is restricted to the $P_{4\text{FD}}$, which are presumed to be under weaker selection, with the trend toward greater GC skew in Arcidae species ($P=0.001$, Fig. 1E) and little difference in AT skew ($P=0.048$, Fig. 1F).

The asymmetric mechanism of mtDNA replication, in which the parental strand is exposed to mutation while it is in a single-stranded state, can account for the strong compositional asymmetry observed in mitogenomes (Reyes *et al.*, 1998). According to this hypothesis, a possible explanation for the marked nucleotide skew in Arcidae species might be that the mitogenome replication in Arcidae is slower than those in other bivalves, exposing the parental strand to deamination for a longer time. Thus, the data of the compositional asymmetry of Arcidae and non-Ar-

Table 3 The bivalves mitochondrial genomes included in the analysis of strand asymmetry in nucleotide composition

Species	Accession no.	mtDNA		P ₁₂₃		P _{4FD}	
		AT skew	GC skew	AT skew	GC skew	AT skew	GC skew
<i>Tegillarca granosa</i>	KJ607173	-0.14	0.41	-0.29	0.43	-0.35	0.72
<i>Anadara vellicata</i>	KP954700	-0.12	0.42	-0.28	0.40	-0.24	0.60
<i>Scapharca kagoshimensis</i>	KF750628	-0.10	0.11	-0.28	0.31	-0.33	0.38
<i>Scapharca broughtonii</i>	AB729113	-0.17	0.36	-0.26	0.33	-0.26	0.43
<i>Trisidos kiyonoi</i>	KU975161	-0.30	0.45	-0.39	0.43	-0.56	0.65
<i>Potiarca pilula</i>	KU975162	-0.15	0.42	-0.27	0.40	-0.25	0.66
<i>Arca navicularis</i>	MG641752	-0.23	0.37	-0.32	0.36	-0.33	0.56
<i>Anadara consociata</i>	MH535977	-0.15	0.46	-0.31	0.47	-0.39	0.73
<i>Barbatia virescens</i>	MF374794	-0.31	0.43	-0.44	0.38	-0.65	0.60
<i>Crassostrea hongkongensis</i>	EU266073	-0.13	0.21	-0.20	0.19	-0.11	0.37
<i>Crassostrea iredalei</i>	FJ841967	-0.10	0.20	-0.20	0.18	-0.10	0.35
<i>Crassostrea sikamea</i>	FJ841966	-0.13	0.21	-0.22	0.19	-0.16	0.33
<i>Crassostrea angulata</i>	FJ841965	-0.13	0.20	-0.22	0.18	-0.16	0.32
<i>Crassostrea ariakensis</i>	FJ841964	-0.13	0.21	-0.22	0.19	-0.15	0.34
<i>Crassostrea virginica</i>	AY905542	-0.13	0.16	-0.22	0.14	-0.14	0.23
<i>Crassostrea nippona</i>	HM015198	-0.10	0.20	-0.20	0.18	-0.08	0.32
<i>Crassostrea gigas</i>	AF177226	-0.13	0.20	-0.22	0.19	-0.15	0.25
<i>Crassostrea gasar</i>	KR856227	-0.13	0.17	-0.22	0.15	-0.13	0.21
<i>Ostrea lurida</i>	KC768038	-0.13	0.18	-0.21	0.18	-0.17	0.28
<i>Ostrea edulis</i>	JF274008	-0.14	0.20	-0.20	0.18	-0.12	0.31
<i>Saccostrea mordax</i>	FJ841968	-0.15	0.21	-0.22	0.19	-0.14	0.40
<i>Saccostrea cucullata</i>	KP967577	-0.15	0.19	-0.22	0.18	-0.15	0.25
<i>Mimachlamys nobilis</i>	FJ415225	-0.25	0.31	-0.34	0.33	-0.39	0.41
<i>Mimachlamys senatoria</i>	KF214684	-0.28	0.32	-0.34	0.33	-0.44	0.40
<i>Placopecten magellanicus</i>	DQ088274	-0.27	0.40	-0.43	0.41	-0.61	0.62
<i>Chlamys farreri</i>	EU715252	-0.18	0.34	-0.31	0.32	-0.31	0.51
<i>Mizuhopecten yessoensis</i>	AB271769	-0.20	0.25	-0.30	0.27	-0.25	0.33
<i>Argopecten irradians</i>	EU023915	-0.26	0.24	-0.35	0.26	-0.38	0.28
<i>Argopecten purpuratus</i>	KF601246	-0.28	0.30	-0.38	0.32	-0.49	0.45
<i>Argopecten ventricosus</i>	KT161261	-0.27	0.28	-0.37	0.30	-0.45	0.38
<i>Pinctada margaritifera</i>	HM467838	-0.23	0.36	-0.32	0.36	-0.37	0.40
<i>Pinctada maxima</i>	GQ452847	-0.31	0.45	-0.39	0.48	-0.55	0.64
<i>Mytilus trossulus</i>	GU936625	-0.09	0.23	-0.16	0.24	0.01	0.45
<i>Mytilus californianus</i>	GQ527172	-0.11	0.26	-0.17	0.27	-0.02	0.50
<i>Mytilus edulis</i>	AY484747	-0.11	0.25	-0.17	0.25	-0.03	0.46
<i>Mytilus galloprovincialis</i>	AY497292	-0.11	0.24	-0.17	0.24	-0.01	0.46
<i>Mytilus coruscus</i>	KJ577549	-0.11	0.26	-0.17	0.26	-0.06	0.49
<i>Brachidontes exustus</i>	KM233636	-0.17	0.20	-0.24	0.19	-0.24	0.18
<i>Perna viridis</i>	JQ970425	-0.19	0.39	-0.26	0.38	-0.24	0.78
<i>Perna perna</i>	KM655841	-0.19	0.20	-0.27	0.22	-0.38	0.25
<i>Musculista senhousia</i>	GU001954	-0.17	0.23	-0.23	0.23	-0.24	0.21
<i>Meretrix petechialis</i>	EU145977	-0.26	0.39	-0.34	0.38	-0.55	0.58
<i>Meretrix lamarckii</i>	GU071281	-0.28	0.43	-0.37	0.39	-0.63	0.65
<i>Meretrix lyrata</i>	KC832317	-0.27	0.45	-0.37	0.40	-0.59	0.60
<i>Meretrix lusoria</i>	GQ903339	-0.26	0.42	-0.35	0.39	-0.57	0.57
<i>Meretrix meretrix</i>	GQ463598	-0.25	0.39	-0.34	0.37	-0.55	0.51
<i>Paphia euglypta</i>	GU269271	-0.15	0.35	-0.22	0.36	-0.16	0.56
<i>Paphia textile</i>	JF969277	-0.14	0.37	-0.25	0.38	-0.21	0.60
<i>Paphia amabilis</i>	JF969276	-0.11	0.29	-0.20	0.30	-0.15	0.42
<i>Paphia undulata</i>	JF969278	-0.15	0.36	-0.25	0.37	-0.16	0.48
<i>Venerupis philippinarum</i>	AB065375	-0.13	0.35	-0.21	0.36	-0.21	0.43
<i>Saxidomus purpuratus</i>	KP419933	-0.22	0.35	-0.31	0.33	-0.35	0.57
<i>Solenaiia oleivora</i>	KF296320	-0.22	0.37	-0.28	0.16	-0.25	0.21
<i>Solen grandis</i>	HQ703012	-0.30	0.39	-0.40	0.41	-0.50	0.59
<i>Solen strictus</i>	JN786377	-0.31	0.37	-0.41	0.39	-0.52	0.51
<i>Solenaiia carinatus</i>	KC848654	-0.22	0.39	-0.27	0.17	-0.20	0.22

(to be continued)

(continued)

Species	Accession no.	mtDNA		P ₁₂₃		P _{4FD}	
		AT skew	GC skew	AT skew	GC skew	AT skew	GC skew
<i>Lucinella divaricata</i>	EF043342	-0.24	0.33	-0.31	0.32	-0.31	0.31
<i>Loripes lacteus</i>	EF043341	-0.23	0.32	-0.31	0.33	-0.31	0.46
<i>Acanthocardia tuberculata</i>	DQ632743	-0.18	0.17	-0.25	0.18	-0.11	0.25
<i>Fulvia mutica</i>	AB809077	-0.13	0.28	-0.24	0.29	-0.16	0.38
<i>Tridacna squamosa</i>	KP205428	-0.12	0.19	-0.19	0.26	-0.05	0.34
<i>Semele scabra</i>	JN398365	-0.23	0.43	-0.32	0.43	-0.36	0.62
<i>Nuttallia olivacea</i>	JN398364	-0.15	0.32	-0.21	0.33	-0.10	0.54
<i>Soletellina diphos</i>	JN398363	-0.26	0.37	-0.33	0.38	-0.45	0.60
<i>Moerella iridescens</i>	JN398362	-0.22	0.35	-0.30	0.32	-0.36	0.34
<i>Solecurtus divaricatus</i>	JN398367	-0.29	0.38	-0.37	0.39	-0.55	0.50
<i>Sinonovacula constricta</i>	EU880278	-0.23	0.36	-0.32	0.37	-0.36	0.63
<i>Coelomacra antiquata</i>	JQ423460	-0.20	0.30	-0.29	0.29	-0.27	0.50
<i>Mactra chinensis</i>	KJ754823	-0.22	0.26	-0.32	0.25	-0.36	0.40
<i>Lutraria rhynchaena</i>	HG799089	-0.28	0.40	-0.37	0.40	-0.47	0.60
<i>Arctica islandica</i>	KF363951	-0.16	0.30	-0.24	0.31	-0.16	0.43
<i>Calypptogena magnifica</i>	KR862368	-0.20	0.39	-0.29	0.39	-0.35	0.64
<i>Mya arenaria</i>	KJ755996	-0.13	0.32	-0.20	0.33	-0.25	0.28
<i>Hiattella arctica</i>	DQ632742	-0.15	0.29	-0.22	0.31	-0.28	0.29
<i>Panopea generosa</i>	KM580067	-0.21	0.38	-0.29	0.39	-0.31	0.33
<i>Panopea globosa</i>	KM580068	-0.27	0.44	-0.35	0.45	-0.38	0.40
<i>Anodonta anatina</i>	KF030964	-0.15	0.30	-0.24	0.18	-0.21	0.20
<i>Anodonta arcaiformis</i>	KF667530	-0.12	0.27	-0.24	0.18	-0.18	0.26
<i>Anodonta lucida</i>	KF667529	-0.13	0.28	-0.23	0.16	-0.19	0.16
<i>Anodonta euscaphys</i>	KP187851	-0.12	0.26	-0.28	0.18	-0.16	0.19
<i>Hyriopsis cumingii</i>	HM347668	-0.23	0.36	-0.30	0.21	-0.27	0.30
<i>Hyriopsis schlegelii</i>	HQ641406	-0.23	0.35	-0.31	0.22	-0.29	0.35
<i>Utterbackia imbecillis</i>	HM856637	-0.15	0.28	-0.23	0.18	-0.20	0.21
<i>Utterbackia peninsularis</i>	HM856635	-0.13	0.26	-0.19	0.13	-0.15	0.18
<i>Unio pictorum</i>	HM014130	-0.19	0.32	-0.25	0.19	-0.25	0.22
<i>Unio douglasiae</i>	KM657954	-0.18	0.32	-0.25	0.19	-0.24	0.21

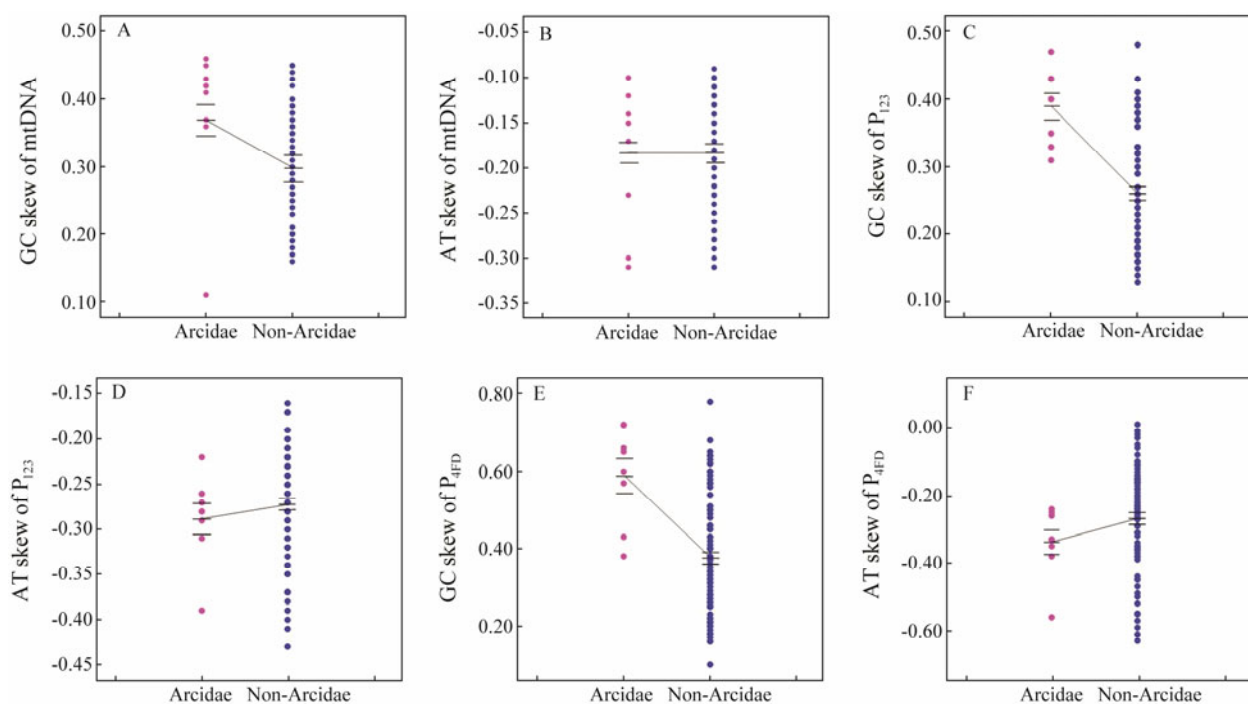


Fig.1 Nucleotide skew values of the whole mtDNA and protein-coding genes of Arcidae and non-Arcidae bivalve mitochondrial genomes. A, GC skews for mtDNA; B, AT skews for mtDNA; C, GC skews for all sites (P₁₂₃); D, AT skews for all sites (P₁₂₃); E, GC skews for the fourfold degenerate sites (P_{4FD}); F, AT skews for the fourfold degenerate sites (P_{4FD}). The average values of each group are indicated along with standard error bars.

cidae group indicated that the presence of the exceptional long no-coding regions may affect replication mechanics. However, the large size of mitogenomes in the Arcidae species does not mean a significant replicative disadvantage. The exceptional long no-coding regions may provide additional replication initiation signals, which can increase the number of genome replicates per template genome (Jiang *et al.*, 2007; Eberhard and Wright, 2016). This is really advantageous if the replication of Arcidae mtDNA is particularly slow, as reflected by the marked nucleotide skew that were found in Arcidae mitogenomes. One way to test this idea is to map replication initiation sites to see whether Arcidae mitogenomes have more replication initiation zones in the non-coding regions.

3.2 Relaxed Selective Constraint on Large Mitogenomes of Arcidae

Previous studies have shown that the mitogenomes are under selection for smaller size, which can cause higher replication and translation efficiency (Rand, 1993). According to this hypothesis, the large mitogenomes of Arcidae species may under different selective constraint compared with small mitogenomes in other bivalves. In order to explore this difference, we assembled a data set of 86 mitogenomes of bivalves and constructed the Maximum Likelihood (ML) phylogenetic tree (Fig.2).

The ratio of nonsynonymous (change in amino acid) and synonymous (silent) substitutions (Ka/Ks) is generally used to measure the selective constraints acting on the protein-coding sequences. The mitochondrial data set of bivalves (listed in Supplementary Table 1) were first divided into 'Arcidae (large mitogenome)' and 'non-Arcidae (small mitogenome)' groups to represent groups with different mitogenome sizes (Fig.2). The Arcidae group has a significant higher mean value of Ka/Ks (0.0705) than the non-Arcidae group (0.0421) ($P=0.002$, Mann-Whitney U-test, Fig.3A). The mean value for Ka is also significantly different between Arcidae and non-Arcidae groups (0.1094 vs. 0.0238, $P<0.001$, Fig.3B), suggesting that the mitochondrial protein-coding genes of Arcidae accumulate more nonsynonymous mutations compared with other bivalves. Considering that the divergences of synonymous mutation rate may bias the results, we compared the average Ks between Arcidae and non-Arcidae groups. The mean value of Ks in Arcidae group (1.8945) is significantly higher than that of the non-Arcidae group (0.7524; $P=0.003$). The greater Ks may result in a smaller Ka/Ks ratio in Arcidae group, making the results more conservative. Therefore, our analyses suggest that the higher Ka/Ks values in Arcidae group are not simply originated from the divergences in synonymous mutation rates.

To identify which mitochondrial protein-coding genes are most affected by the selective constraints, we tested the Ka/Ks ratio for each of the 12 mitochondrial genes (Table 1). Seven protein-coding genes (*atp6*, *cox1-3*, *nad1*, *nad4* and *nad5*) show significantly higher Ka/Ks ratios in Arcidae species (Fig.4). This result suggests that these genes may have experienced more relaxed functional con-

straints.

In order to determine whether the Ka/Ks variations depend upon the mitogenome size, or they just reflect a general pattern of molecular evolution for bivalves, we repeated the above analysis for histone H3, a nuclear gene from 49 bivalves, which is independent of mitogenome size (listed in Table 2). However, the Ka/Ks ratio of histone H3 gene is not significantly different between the two groups (0.0001 vs. 0.0085, $P=0.142$).

An alternative hypothesis to explain this finding is that the selective constraints are relaxed on the large mitogenomes of Arcidae species. Because mitochondria play a crucial role in energy generation, mitochondrial genes are more sensitive to the energy-related selective pressures. Higher rates of nonsynonymous substitutions in mtDNA genes may lead to more radical amino acid substitutions (Hanada *et al.*, 2007), resulting a reduction in electron-transferring respiratory chain activity (Weber *et al.*, 1997; Brown *et al.*, 2000). Previous study has showed that low metabolic rates is correlated with relaxed selective constraints on mitochondrial genes (Chong and Mueller, 2012). Based on this hypothesis, Arcidae species may be more likely to survive and reproduce with lower metabolic requirements than other bivalves under similar environment. This coincides with the biological characteristics of Arcidae species. Some Arcidae species are more tolerant to asphyxiation as they can more economically consume oxygen, such as the arcid clam *Scapharca kagoshimensis*, which can adapt well to oxygen content change in the water (Anistratenko and Khaliman, 2006; Soldatov *et al.*, 2009). When the organisms expose to environmental hypoxia, the energy production in mitochondria is slowed, and metabolic rate will be suppressed (Richards, 2011). We thus deduced that the relaxation of selective constraint on large mitogenomes of arcid species is related to their low metabolic rates. The relaxation of selective constraints contributes to generate 'new' (adapted) mitochondrial genes, and positive selection is the basis of adaptive evolution (Shen *et al.*, 2010). Thus the positive selection may have occurred on some mitochondrial genes in Arcidae species to generate the adapted genes.

4 Conclusions

In the present study, we conducted a comparative analysis of 86 bivalve mitogenomes (including 9 arcid mitogenomes) to explore the differences of base composition and selective constraints between the large mitogenomes in Arcidae and small ones in other bivalves. Arcidae mitogenomes have significantly greater GC skews in their coding sequences. The mitochondrial protein-coding genes of Arcidae species had significant higher Ka/Ks than other bivalves. Seven protein-coding genes (*atp6*, *cox1-3*, *nad1*, *nad4* and *nad5*) are most affected by the selective constraints. These divergences are not observed in the nuclear gene (histone H3). The replication of the large mitogenomes in Arcidae may be slower than those in other bivalves. The large mitogenomes of Arcidae experienced more relaxed selective constraints, which is supposed to

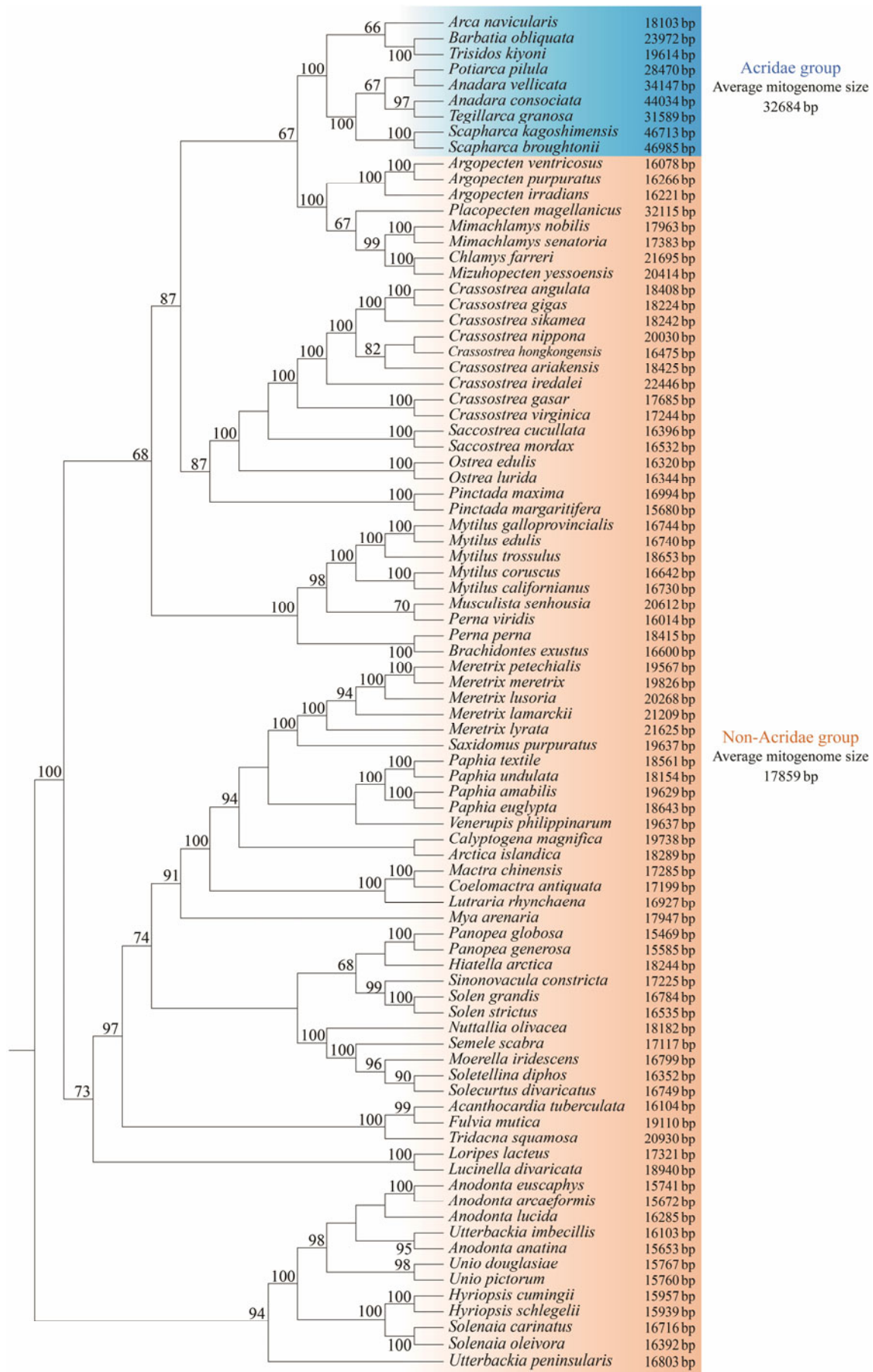


Fig.2 Bivalve phylogenetic tree constructed from 12 mitochondrial protein-coding genes with the Maximum Likelihood method. Arcidae species are marked in blue. The mitogenome size of each bivalve species are indicated. The average mitogenome size of Arcidae and non-Arcidae groups are presented.

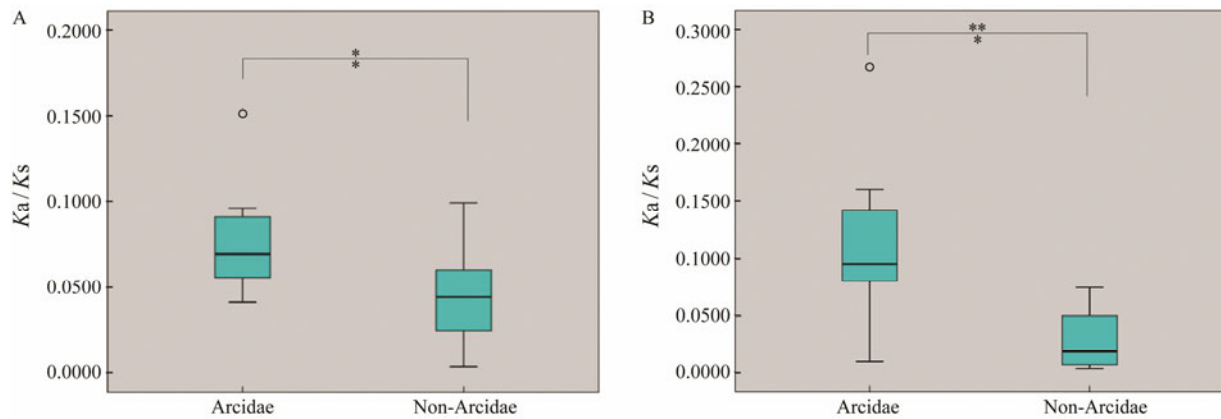


Fig.3 Comparisons of K_a/K_s ratios (A) and K_a (B) between Arcidae and non-Arcidae groups. * $0.01 < P < 0.05$, ** $0.001 < P < 0.01$, *** $P < 0.001$.

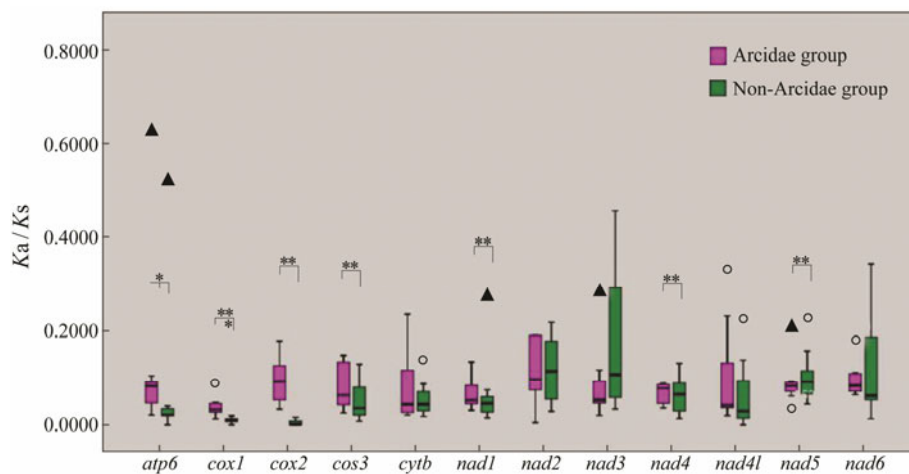


Fig.4 Comparisons of K_a/K_s ratios of the 12 mitochondrial protein-coding genes between Arcidae and non-Arcidae groups. * $0.01 < P < 0.05$, ** $0.001 < P < 0.01$, *** $P < 0.001$.

be related to their low metabolic rates, a response to hypoxia exposure.

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